

Original Article

Identification and bioinformatic analysis of *invA* gene of *Salmonella* in free range chicken

Identificação e análise bioinformática do gene *invA* de *Salmonella* em frango caipira

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Abstract

Salmonella is a serious cause of the health issues in human and animal worldwide. *Salmonella* has been isolated from different biological samples and it considers as the key role in induction of inflammation of gastrointestinal tract which in turn cause diarrhoea in different species. To further understand the involvement of *Salmonella* in contaminating and infecting fresh eggs and meat of free-range chicken. This study aimed to establish the microbiological and molecular detections of *Salmonella* in the cloaca of the free-range chicken and to identify predicted biological functions using Kyoto Encyclopedia of Gene and Genomic (KEGG) pathways and protein-protein interaction. Cloacal swabs were collected from free range chicken raised in the local farm in Duhok city. The isolates were cultured and biochemical test performed using XLD and TSI, respectively. Molecular detection and functional annotation of *invA* gene was carried out using Conventional PCR and bioinformatics approaches. The present study found that *Salmonella* was detected in 36 out of 86 samples using microbiological methods. To confirm these findings, *invA* gene was utilised and 9 out of 36 *Salmonella* isolates have shown a positive signal of *invA* by agarose gel. In addition, bioinformatic analysis revealed that *invA* gene was mainly associated with bacterial secretion processes as well as their KEGG terms and Protein-Protein Interaction were involved in bacterial invasion and secretion pathways. These findings suggested that *invA* gene plays important role in regulating colonization and invasion processes of *Salmonella* within the gut host in the free range chicken.

Keywords: *invA*, *Salmonella*, free range chicken, conventional PCR.

Resumo

A salmonela é uma causa séria de problemas de saúde em humanos e animais em todo o mundo. *Salmonella* tem sido isolada de diferentes amostras biológicas e é considerada como o principal papel na indução da inflamação do trato gastrointestinal que por sua vez causa diarreia em diferentes espécies. Compreender melhor o envolvimento da *Salmonella* na contaminação e infecção de ovos frescos e carne de frango caipira. Este estudo teve como objetivo estabelecer as detecções microbiológicas e moleculares de *Salmonella* na cloaca da galinha caipira e identificar as funções biológicas previstas usando as vias da Enciclopédia de Gene e Genômica de Kyoto (KEGG) e interação proteína-proteína. *Suabes cloacais* foram coletados de galinhas criadas ao ar livre em fazenda local na cidade de Duhok. Os isolados foram cultivados e o teste bioquímico realizado com XLD e TSI, respectivamente. A detecção molecular e anotação funcional do gene *invA* foi realizada usando PCR convencional e abordagens de bioinformática. O presente estudo descobriu que *Salmonella* foi detectada em 36 das 86 amostras usando métodos microbiológicos. Para confirmar esses achados, o gene *invA* foi utilizado e 9 dos 36 isolados de *Salmonella* mostraram um sinal positivo de *invA* pelo gel de agarose. Além disso, a análise bioinformática revelou que o gene *invA* estava associado principalmente a processos de secreção bacteriana, assim como seus termos KEGG e interação proteína-proteína estavam envolvidos na invasão bacteriana e nas vias de secreção. Esses achados sugeriram que o gene *invA* desempenha um papel importante na regulação dos processos de colonização e invasão de *Salmonella* dentro do hospedeiro intestinal na galinha caipira.

Palavras-chave: *invA*, *Salmonella*, galinha caipira, PCR convencional.

1. Introduction

Salmonella is a significant cause of poor health in human and animal worldwide. Studies indicate that more than 90 million cases of foodborne disease caused by *Salmonella enterica* subsp. *enterica* serovar Typhimurium. Food such as poultry, eggs and dairy products are

considered as the major sources of *Salmonella* infections (Balasubramanian et al., 2019). The main symptom of *Salmonella* infection is gastroenteritis, however, it can cause typhoid fever, and paratyphoid fever or even cause death in serious infections in human (Eng et al., 2015).

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Salmonella enterica is an enteropathogenic flagellated rod-shaped Gram-negative bacilli that is considered as a key cause of enteric illness worldwide (Gal-Mor et al., 2014; Balasubramanian et al., 2019). The species *S. enterica* is divided into six subspecies: *enterica*, *salamae*, *arizonae*, *diarizonae*, *houtenae*, and *indica* and each of them has many serotypes or serovars (Kirk et al., 2015). *Salmonella enterica* serotype Typhimurium (*S. Typhimurium*) and *Salmonella enterica* serotype Enteritidis (*S. Enteritidis*) are the most common cause of disease in humans and veterinary animals in the world. Poultry and its products are the main sources of infection with *Salmonella* and can easily transmit *salmonella* from reservoirs to other animals and humans (Kirk et al., 2015; Besharati et al., 2020; Vaez et al., 2020). Poultry infected with *S. pullorum* and *gallinarum* reveals symptoms of white diarrhoea and fowl typhoid. Early detection of *salmonella* has been used to improve the poultry reproductive efficiency in order to meet a rapid rise in global requirements for meat and egg products. However, this can be affected by several factors including the presence of known virulence factors that have been associated with severe human disease in *salmonella* species. Recently, free-range chicken farms have become major contributors to organic food production in several Asian countries including Iraq due to health and economic benefits (Rossi et al., 2018). The young chickens usually obtained from local commercial poultry hatchery or can be hatched under hen, afterward they are raised without using antibiotics or growth hormones for therapy or enhancing growth compared with chickens raised in commercial broiler farms (Zhao et al., 2016). Free-range chickens in developing countries are very popular because they can easily access green spaces, consume leftovers and use less antibiotics. In addition, free-range chickens can provide a side income and a good source of meat and eggs (Padhi, 2016). However, free-range chickens grow very slowly and are more likely prone to diseases including gastroenteritis that is caused by *Salmonella* spp. (Jafari et al., 2007; Sibanda et al., 2020). Improvement in poultry production will be achieved by better understanding the molecular mechanism that underlies the *Salmonella* pathogenicity as a result of stimulation of the several virulence factors of the *salmonella* strains. Several groups have developed different molecular techniques and bioinformatic software to study the genome of *Salmonella* (Shi et al., 2015; Diep et al., 2019). Moreover, studies have revealed that several invasion genes such as a subset of *Inv* genes can control the invasion and colonisation of *Salmonella enterica* and *typhimurium* in the gut wall of young aged chick and adult chicken (Harvey et al., 2011). In addition, several virulence genes have been involved in colonisation in the gut system of different species, including *Salmonella* pathogenicity islands (SPI1 and SPI2) (Ilyas et al., 2017). This study aimed to detect the *salmonella* isolated from the chicken cloaca using microbiological and molecular approaches as well as provide an insight into the functional roles of *invA* gene in invasion and colonisation of *Salmonella* spp.

2. Materials and Methods

2.1. Ethics statement

Ethical approval was given by the ethics committee at the college of Veterinary Medicine, university of Duhok (DR2020820CV) for all experimental procedures.

2.2. Sample collection

86 fresh cloacal swabs were obtained from three free-range chicken farms in Duhok city, Iraq. Fresh samples were collected using sterile cotton swabs. After sampling, the swabs were transported to Duhok research centre at the College of Veterinary Medicine, University of Duhok and processed for microbiological and molecular studies.

2.3. Isolation of *Salmonella* isolates

The cotton swabs were immediately put into vials containing 10 mL of warm buffered peptone water for non-selective pre-enrichment, shaken sufficiently and incubated overnight with swab at 37 °C. Then 0.1 mL of the enrichment broth was added to Rappaport-Vassiliadis Medium at 42 °C for 24 h. Afterward, a 10 µL bacteriological loop was used to streak the RVS culture on to Xylose Lysine Deoxycholate agar and incubated at 37 °C ± 1 overnight. Suspected *Salmonella* colonies were then sub-cultured to Xylose Lysine Deoxycholate (XLD) and incubated at 37 °C ± 1 for 24 h (Park et al., 2012). Suspected *Salmonella* isolates were maintained in 25% glycerol and brain heart infusion broth (BHIB) and stored at -20 °C for further processing.

2.4. Biochemical characterization of isolates

The *Salmonella* isolates were subjected to the Triple sugar iron (TSI) agar test. The test was performed according to the standard protocols. A pure colony was stab-inoculated into the butt of the TSI medium and later streaked on the slant and incubated aerobically at 37 °C for 24 hours (Waltman et al., 2008). *Salmonella*-positive colonies were collected and kept under -20 °C for further use.

2.5. DNA extraction

Two to three colonies were collected from positive *Salmonella* Xylose Lysine Deoxycholate agar plates, placed in a microfuge tube and resuspended in 100 µL of sterile water. The suspension was incubated at 100 °C for 10 min. The bacterial debris was removed by centrifugation (9600 g, 10 min) and supernatant was transferred into a new tube, followed by incubating samples on ice for five minutes then stored at -20 °C until ready for use.

2.6. PCR characterisation of *invA* gene

A PCR was used to identify the isolates collected during sampling. Isolates were identified as *Salmonella* through the amplification of an *InvA* gene (primer pairs 5'-ACAGTGCTCGTTTACGACCTGAAT-3' and 5'-AGACGACTGGTACTGATCGATAAT-3') (Bhatta et al., 2007). The primers were blasted against the nucleotide database of the EBI (BLASTN, version 2.9.0+) website to ensure identity among reported BLAST sequences for

the target gene and the absence of significant homology with other microorganism sequences (EMBL-EBI, 2022). PCR reactions were conducted in a total volume of 20 μ L. Each reaction contained 10 μ L 2 \times Ruby PCR Master (Ruby Taq Master, Jena Bioscience, Germany), 0.3 μ M of each forward and reverse primer for InvA and 2 μ L DNA. PCR cycling conditions were as follows; 1 min at 94 $^{\circ}$ C followed by 30 cycles of 30 s at 94 $^{\circ}$ C, 30s at 60 $^{\circ}$ C and 2 min at 72 $^{\circ}$ C followed by 10 min at 72 $^{\circ}$ C (Chiu and Ou, 1996).

2.7. Gel electrophoresis of amplicons

Detection of invA amplicons was carried out using a 1.5% (w/v) agarose gel prepared from 1X tris acetate ethylenediamineacetate (TAE) buffer with Prime Safe Dye (GeNet Bio, Korea). A DNA marker (Jena bioscience, Germany) of was used. The gel was allowed to run at 85 volts, 300 amperes for 45 mins in 1X TAE buffer and was visualised under the UV transilluminator light.

2.8. Gene enrichment analysis

ClueGO (v2.5.7) software was used to detect Gene Ontology (GO) term and Kyoto Encyclopedia of Gene and Genome (KEGG) pathways. ClueGO enable analysis of genes from several organism including salmonella and it can recognise many identifier types based on information from different sources such as NCBI, UniPrkB and Ensembl (Bindea et al., 2009). Two-sided hypergeometric test used for enrichment analyses, Benjamini–Hochberg correction was utilized for P value correction and Kappa coefficient of 0.4 was used to detect the similarities of GO terms for associated genes. The resulting GO terms with $P < 0.01$ and KEGG pathways with $P < 0.05$ were considered significant. Furthermore, the results were constructed and visualized by Cytoscape (v3.8.2)

2.9. Protein–protein interactions network

Protein–protein interaction analysis of the miRNA target genes was performed using STRING (2022) based on Salmonella species. STRING is an online tool designed to determine the potential predicted and experimental

information of interaction between proteins. The interactions between the invA gene and associated proteins were mapped into PPIs network with confidence score of >0.7 as the cutoff standard. Cytoscape–plugin known as network analyzer (v4.4.6) was utilized to construct the score of gene nodes.

3. Results

3.1. Identification of Salmonella using microbiological methods

Out of 86 isolates, 36 were salmonella positive based on their characteristics of colony morphological and biochemical properties. Salmonella was identified by typical pink with black centers colonies on the XLD agar and can be easily differentiated from other organisms (Figure 1A). Confirmatory test was also carried out on the Salmonella spp. using TSI which measures the precipitation of hydrogen sulfide and utilization of glucose, lactose and sucrose. Salmonella isolates found to be TSI positive, the slant showed red color of the medium, the butt was yellow due to glucose fermentation as well as the TSI test revealed blackening of medium as a result of H₂S production (Figure 1B).

3.2. Identification of Salmonella using conventional PCR

The goal of this study was to elucidate the detection of the invA gene of Salmonella in the cloacal samples of the free-range chicken. The InvA primer used in this study was designated by Bhatta et al group is 244-bp long and located on chromosome I of Salmonella spp. Using EBI-BLASTN+, Blast alignment analysis revealed that this sequence was 100% similar with several Salmonella species such as Salmonella gallinarum, Salmonella enterica subsp. enterica serovar Typhimurium, paratyphi, enteritidis and pullorum (Table S1). PCR amplification of positive invA gene was observed by agarose gel (Figure 2). Among of the 36 representative isolates, about 9 isolates showed positive amplification of the invA gene while about 27 isolates were negative.

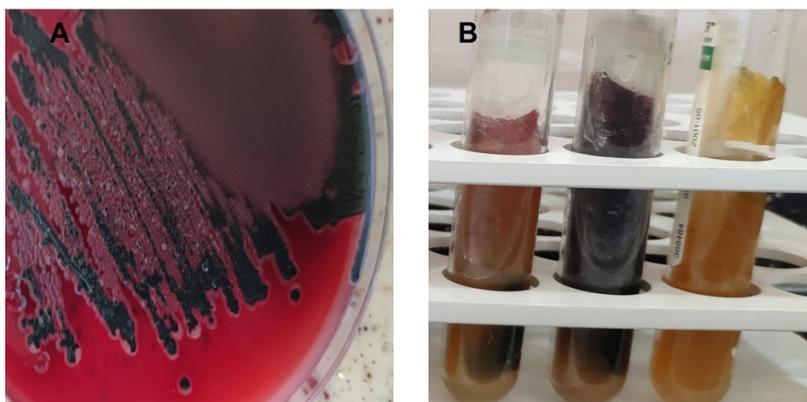


Figure 1. A representative schematic of the (A) Phenotypic characteristics of Salmonella spp. on the XLD agar; (B) TSI test showed positive production of H₂S and fermentation of glucose tube 1 positive (Alkaline/Acid, -Gas, + H₂S), tube 2 positive (Alkaline/Acid, -Gas, + H₂S), tube 3 negative (Acid/Acid).

3.3. Gene enrichment functional analysis

Gene Ontology (GO) enrichment analysis and Kyoto Encyclopedia of Genes and Genomics (KEGG) pathway analysis showed that *invA* was significantly enriched for 6 biological process (BP) and 3 cellular components (CC). The most significant GO terms were secretion by cell (GO:0032940) and plasma membrane (GO:0005886) in BP and CC categories, respectively (Figure 3). KEGG enrichment showed significantly enriched pathways for *invA* gene, including bacterial invasion and bacterial secretion pathways (sty05100, sty03070), respectively (Figures 4 and 5).

3.4. PPI network construction and network analysis of target genes

Protein interactions were explored and analyzed using STRING. As shown in Figure 6A, a total of 1635 edges and 91 nodes were obtained, Using the network analyzer in Cytoscape, we identified top core genes with the highest node degrees (*spaO*, *invF*, *prgH*, *sipB*, *invE*, *invG*, *spaI*, *spaS*, *sptP*, *sspH*, *sicA*, *prgK*, *envE*, *spaP*, *spaN*, *sopB*, *sopD*) (Figure 6B). These results showed a significant enrichment of genes for GO terms such as peptide secretion

(GO:000279). Top KEGG pathways significantly associated with genes in the module were Bacterial invasion of epithelial cells (KEGG:05100) and Salmonella infection (KEGG:05132).

4. Discussion

Salmonella considers as major cause of food born disease in human and animals worldwide. Backyard or free-range chicken have been widely grown by individuals as a source of fresh nutritious eggs and meat, however, free-range chicken is more susceptible to transmit Salmonella and induce infection among species (Ghoddusi et al., 2015). The conventional microbiological methods, including culture using selective medium such as XLD and biochemical test such as TSI have shown that 36 out of 86 cloacal samples were identified to be Salmonella spp. However, due to low specificity of these techniques (Sousa and Pereira, 2013), these results were validated using *invA* gene as key marker for detection of Salmonella. *invA* gene is highly conserved across Salmonella species, located in Salmonella pathogenicity island-1 and it is critically involved in regulating pathogenicity and

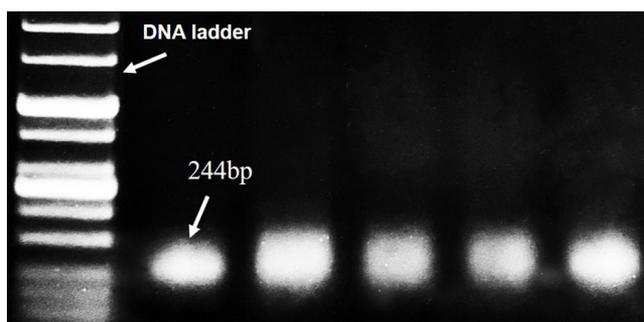


Figure 2. Analysis of PCR amplified *invA* gene on the 1.5% agarose gel electrophoresis. Lane 1 is a 100 bp DNA marker), other lanes (244 bp) are the amplified fragment for *invA* gene.

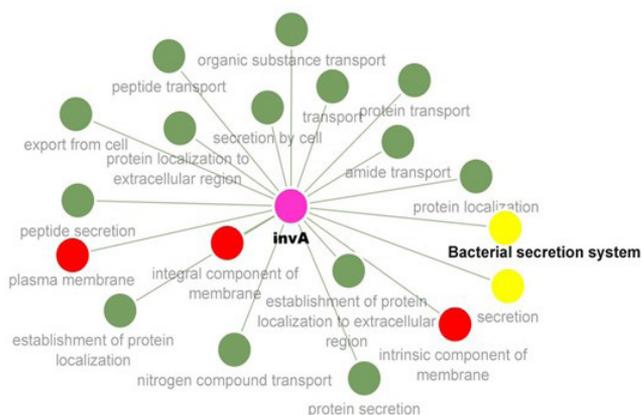


Figure 3. Functional GO terms and KEGG pathway enriched for *invA* gene using ClueGO. The node represents a GO term, each color represents a functional group and the node size represents the term enrichment significance, $p \leq 0.01$. A. Enriched biological processes (BP) (green), Cellular components (CC) (red colour) and KEGG pathway (Yellow).

invA, invF, invE, invG Invasion proteins
 sipB Cell invasion protein SipB
 sptP protein tyrosine phosphatase
 sspH Salmonella secreted protein H1
 sicA Salmonella invasion chaperone
 prgK, prgH Lipoprotein Prg
 envE Probable lipoprotein EnvE
 spaP, spaN, spaO, spaI, spaS Surface presentation of antigens protein
 hilA hyperinvasive locus A
 sopB, sopD secreted protein in the Sop family
 spp Species
 TSI Triple sugar iron
 XLD Xylose Lysine Deoxycholate
 RV Rappaport-Vassiliadis Medium

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References

- AKINOLA, S.A., MWANZA, M. and ATEBA, C.N., 2019. Occurrence, genetic diversities and antibiotic resistance profiles of *Salmonella* serovars isolated from chickens. *Infection and Drug Resistance*, vol. 12, pp. 3327-3342. <http://dx.doi.org/10.2147/IDR.S217421>. PMID:31695452.
- AL-IEDANI, A.A., MUSTAFA, J.Y. and HADI, N.S., 2015. Comparison of techniques used in isolation and identification of salmonella spp. and related genera from raw meat and abattoir environment of Basrah. *International Journal for Sciences and Technology*, vol. 143, no. 3101, pp. 1-7. <http://dx.doi.org/10.12816/0024585>.
- BALASUBRAMANIAN, R., IM, J., LEE, J.S., JEON, H.J., MOGENI, O.D., KIM, J.H., RAKOTOZANDRINDRAINY, R., BAKER, S. and MARKS, F., 2019. The global burden and epidemiology of invasive non-typhoidal *Salmonella* infections. *Human Vaccines & Immunotherapeutics*, vol. 15, no. 6, pp. 1421-1426. <http://dx.doi.org/10.1080/21645515.2018.1504717>. PMID:30081708.
- BESHARATI, S., SADEGHI, A., AHMADI, F., TAJEDDIN, E., MOHAMMAD SALEHI, R., FANI, F., POULADFAR, G., NIKMANESH, B., MAJIDPOUR, A., SOLEYMANZADEH MOGHADAM, S., MIRAB SAMIEE, S., RAHNAMAYE FARZAMI, M., RAHBAR, M., ESLAMI, P., RAKHSHANI, N., ESHRATI, B., GOUYA, M.M., FALLAH, F., KARIMI, A., OWLIA, P. and ALEBOUYEH, M., 2020. Serogroups, and drug resistance of nontyphoidal *Salmonella* in symptomatic patients with community-acquired diarrhea and chicken meat samples in Tehran. *Iranian Journal of Veterinary Research*, vol. 21, no. 4, pp. 269-278. <http://dx.doi.org/10.22099/ijvr.2020.36912.5387>. PMID:33584839.
- BHATTA, D.R., BANGTRAKULNONT, A., TISHYADHIGAMA, P., SAROJ, S.D., BANDEKAR, J.R., HENDRIKSEN, R.S. and KAPADNIS, B.P., 2007. Serotyping, PCR, phage-typing and antibiotic sensitivity testing of *Salmonella* serovars isolated from urban drinking water supply systems of Nepal. *Letters in Applied Microbiology*, vol. 44, no. 6, pp. 588-594. <http://dx.doi.org/10.1111/j.1472-765X.2007.02133.x>. PMID:17576218.
- BINDEA, G., MLECNIK, B., HACKL, H., CHAROENTONG, P., TOSOLINI, M., KIRILOVSKY, A., FRIDMAN, W.H., PAGÈS, F., TRAJANOSKI, Z. and GALON, J., 2009. ClueGO: a Cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks. *Bioinformatics*, vol. 25, no. 8, pp. 1091-1093. <http://dx.doi.org/10.1093/bioinformatics/btp101>. PMID:19237447.
- CHIU, C.H. and OU, J.T., 1996. Rapid identification of *Salmonella* serovars in feces by specific detection of virulence genes, invA and spvC, by an enrichment broth culture-multiplex PCR combination assay. *Journal of Clinical Microbiology*, vol. 34, no. 10, pp. 2619-2622. <http://dx.doi.org/10.1128/jcm.34.10.2619-2622.1996>. PMID:8880536.
- COLLAZO, C.M. and GALÁN, J.E., 1997. The invasion-associated type III system of *Salmonella typhimurium* directs the translocation of Sip proteins into the host cell. *Molecular Microbiology*, vol. 24, no. 4, pp. 747-756. <http://dx.doi.org/10.1046/j.1365-2958.1997.3781740.x>. PMID:9194702.
- CORNELIS, G.R., 2006. The type III secretion injectisome. *Nature Reviews. Microbiology*, vol. 4, no. 11, pp. 811-825. <http://dx.doi.org/10.1038/nrmicro1526>. PMID:17041629.
- DARWIN, K.H. and MILLER, V.L., 1999. Molecular basis of the interaction of *Salmonella* with the intestinal mucosa. *Clinical Microbiology Reviews*, vol. 12, no. 3, pp. 405-428. <http://dx.doi.org/10.1128/CMR.12.3.405>. PMID:10398673.
- DIEP, B., BARRETTO, C., PORTMANN, A.C., FOURNIER, C., KARCZMAREK, A., VOETS, G., LI, S., DENG, X. and KLIJN, A., 2019. *Salmonella* Serotyping; Comparison of the Traditional Method to a Microarray-Based Method and an in silico Platform Using Whole Genome Sequencing Data. *Frontiers in Microbiology*, vol. 10, pp. 2554. <http://dx.doi.org/10.3389/fmicb.2019.02554>. PMID:31781065.
- EL-SHARKAWY, H., TAHOUN, A., EL-GOHARY, A.E.A., EL-ABASY, M., EL-KHAYAT, F., GILLESPIE, T., KITADE, Y., HAFEZ, H.M., NEUBAUER, H. and EL-ADAWY, H., 2017. Epidemiological, molecular characterization and antibiotic resistance of *Salmonella enterica* serovars isolated from chicken farms in Egypt. *Gut Pathogens*, vol. 9, no. 1, pp. 8. <http://dx.doi.org/10.1186/s13099-017-0157-1>. PMID:28203289.
- EMBL-EBI [online], 2022 [viewed 23 April 2022]. Available from: <https://www.ebi.ac.uk/Tools/sss/ncbiblast/nucleotide.html>
- ENG, S.K., PUSPARAJAH, P., AB MUTALIB, N.-S., SER, H.-L., CHAN, K.-G. and LEE, L.-H., 2015. *Salmonella*: a review on pathogenesis, epidemiology and antibiotic resistance. *Frontiers in Life Science*, vol. 8, no. 3, pp. 284-293. <http://dx.doi.org/10.1080/21553769.2015.1051243>.
- GALÁN, J.E. and WOLF-WATZ, H., 2006. Protein delivery into eukaryotic cells by type III secretion machines. *Nature*, vol. 444, no. 7119, pp. 567-573. <http://dx.doi.org/10.1038/nature05272>. PMID:17136086.
- GAL-MOR, O., BOYLE, E.C. and GRASSL, G.A., 2014. Same species, different diseases: how and why typhoidal and non-typhoidal *Salmonella enterica* serovars differ. *Frontiers in Microbiology*, vol. 5, pp. 391. <http://dx.doi.org/10.3389/fmicb.2014.00391>. PMID:25136336.
- GENOMENET, 2022a [viewed 23 April 2022]. *Bacterial invasion of epithelial cells* [online]. Available from: https://www.genome.jp/kegg-bin/show_pathway?sty05100
- GENOMENET, 2022b [viewed 23 April 2022]. *Bacterial secretion system* [online]. Available from: <https://www.genome.jp/pathway/sty03070>
- GHODDUSI, A., NAYERI FASAEI, B., KARIMI, V., ASHRAFI TAMAI, I., MOULANA, Z. and ZAHRAEI SALEHI, T., 2015. Molecular identification of *Salmonella* Infantis isolated from backyard

- chickens and detection of their resistance genes by PCR. *Iranian Journal of Veterinary Research*, vol. 16, no. 3, pp. 293-297. PMID:27175192.
- GUO, X., CHEN, J., BEUCHAT, L.R. and BRACKETT, R.E., 2000. PCR detection of *Salmonella enterica* serotype Montevideo in and on raw tomatoes using primers derived from *hilA*. *Applied and Environmental Microbiology*, vol. 66, no. 12, pp. 5248-5252. <http://dx.doi.org/10.1128/AEM.66.12.5248-5252.2000>. PMID:11097898.
- HARVEY, P.C., WATSON, M., HULME, S., JONES, M.A., LOVELL, M., BERTCHIERI JUNIOR, A., YOUNG, J., BUMSTEAD, N. and BARROW, P., 2011. *Salmonella enterica* serovar typhimurium colonizing the lumen of the chicken intestine grows slowly and upregulates a unique set of virulence and metabolism genes. *Infection and Immunity*, vol. 79, no. 10, pp. 4105-4121. <http://dx.doi.org/10.1128/IAI.01390-10>. PMID:21768276.
- ILYAS, B., TSAI, C.N. and COOMBES, B.K., 2017. Evolution of *Salmonella*-host cell interactions through a dynamic bacterial genome. *Frontiers in Cellular and Infection Microbiology*, vol. 7, pp. 428. <http://dx.doi.org/10.3389/fcimb.2017.00428>. PMID:29034217.
- JAFARI, R.A., GHORBANPOU, M. and JAIDERI, A., 2007. An investigation into *Salmonella* infection status in backyard chickens in Iran. *International Journal of Poultry Science*, vol. 6, no. 3, pp. 227-229. <http://dx.doi.org/10.3923/ijps.2007.227.229>.
- JAJERE, S.M., HASSAN, L., ABDUL AZIZ, S., ZAKARIA, Z., ABU, J., NORDIN, F. and FAIZ, N.M., 2019. *Salmonella* in native "village" chickens (*Gallus domesticus*): prevalence and risk factors from farms in South-Central Peninsular Malaysia. *Poultry Science*, vol. 98, no. 11, pp. 5961-5970. <http://dx.doi.org/10.3382/ps/pez392>. PMID:31392329.
- KIM, S.I., KIM, S., KIM, E., HWANG, S.Y. and YOON, H., 2018. Secretion of *Salmonella* pathogenicity island 1-encoded type III secretion system effectors by outer membrane vesicles in *Salmonella enterica* serovar Typhimurium. *Frontiers in Microbiology*, vol. 9, pp. 2810. <http://dx.doi.org/10.3389/fmicb.2018.02810>. PMID:30532744.
- KIRK, M.D., PIRES, S.M., BLACK, R.E., CAIPO, M., CRUMP, J.A., DEVLEESSCHAUWER, B., DÖPFER, D., FAZIL, A., FISCHER-WALKER, C.L., HALD, T., HALL, A.J., KEDDY, K.H., LAKE, R.J., LANATA, C.F., TORGERSON, P.R., HAVELAAR, A.H. and ANGULO, F.J., 2015. World Health Organization estimates of the global and regional disease burden of 22 foodborne bacterial, protozoal, and viral diseases, 2010: a data synthesis. *PLoS Medicine*, vol. 12, no. 12, pp. e1001921-e1001921. <http://dx.doi.org/10.1371/journal.pmed.1001921>. PMID:26633831.
- LARA-TEJERO, M. and GALÁN, J.E., 2009. *Salmonella enterica* serovar typhimurium pathogenicity island 1-encoded type III secretion system translocases mediate intimate attachment to nonphagocytic cells. *Infection and Immunity*, vol. 77, no. 7, pp. 2635-2642. <http://dx.doi.org/10.1128/IAI.00077-09>. PMID:19364837.
- LARA-TEJERO, M., KATO, J., WAGNER, S., LIU, X. and GALÁN, J.E., 2011. A sorting platform determines the order of protein secretion in bacterial type III systems. *Science*, vol. 331, no. 6021, pp. 1188-1191. <http://dx.doi.org/10.1126/science.1201476>. PMID:21292939.
- LARA-TEJERO, M., QIN, Z., HU, B., BUTAN, C., LIU, J. and GALÁN, J.E., 2019. Role of SpaO in the assembly of the sorting platform of a *Salmonella* type III secretion system. *PLoS Pathogens*, vol. 15, no. 1, e1007565. <http://dx.doi.org/10.1371/journal.ppat.1007565>. PMID:30668610.
- LOU, L., ZHANG, P., PIAO, R. and WANG, Y., 2019. *Salmonella* pathogenicity island 1 (SPI-1) and its complex regulatory network. *Frontiers in Cellular and Infection Microbiology*, vol. 9, pp. 270. <http://dx.doi.org/10.3389/fcimb.2019.00270>. PMID:31428589.
- MKANGARA, M., MBEGA, E.R. and CHACHA, M., 2020. Molecular identification of *Salmonella* Typhimurium from village chickens based on *invA* and *spvC* genes. *Veterinary World*, vol. 13, no. 4, pp. 764-767. <http://dx.doi.org/10.14202/vetworld.2020.764-767>.
- MONJARÁS FERIA, J.V., LEFEBRE, M.D., STIERHOF, Y.D., GALÁN, J.E. and WAGNER, S., 2015. Role of autocleavage in the function of a type III secretion specificity switch protein in *Salmonella enterica* serovar Typhimurium. *mBio*, vol. 6, no. 5, e01459-15. <http://dx.doi.org/10.1128/mBio.01459-15>. PMID:26463164.
- PADHI, M.K., 2016. Importance of indigenous breeds of chicken for rural economy and their improvements for higher production performance. *Scientifica*, vol. 2016, pp. 2604685. <http://dx.doi.org/10.1155/2016/2604685>. PMID:27144053.
- PARK, S.-H., RYU, S. and KANG, D.-H., 2012. Development of an improved selective and differential medium for isolation of *Salmonella* spp. *Journal of Clinical Microbiology*, vol. 50, no. 10, pp. 3222-3226. <http://dx.doi.org/10.1128/JCM.01228-12>. PMID:22814469.
- PORTA, C., PAGLINO, C. and MOSCA, A., 2014. Targeting PI3K/Akt/mTOR signaling in cancer. *Frontiers in Oncology*, vol. 4, pp. 64. <http://dx.doi.org/10.3389/fonc.2014.00064>. PMID:24782981.
- RAFFATELLU, M., WILSON, R.P., CHESSA, D., ANDREWS-POLYMENIS, H., TRAN, Q.T., LAWHON, S., KHARE, S., ADAMS, L.G. and BÄUMLER, A.J., 2005. SipA, SopA, SopB, SopD, and SopE2 contribute to *Salmonella enterica* serotype typhimurium invasion of epithelial cells. *Infection and Immunity*, vol. 73, no. 1, pp. 146-154. <http://dx.doi.org/10.1128/IAI.73.1.146-154.2005>. PMID:15618149.
- ROSSI, G., CONTI, L., BAMBI, G., MONTI, M. and BARBARI, M., 2018. Poultry farming solutions for a sustainable development of marshlands areas of South Iraq. *Agronomy Research*, vol. 16, no. 2, pp. 574581.
- SAFARPOUR-DEHKORDI, M., DOOSTI, A., JAMI, M.S. and GHOLIPOUR, A., 2018. Quantitative Real Time PCR study on the effects of *hilA* gene deletion on the expression of pathogenicity genes in *Salmonella enterica* ssp. *Journal of BioScience and Biotechnology*, vol. 7, no. 2-3, pp. 73-78.
- SCHOLZ, H.C., ARNOLD, T., MARG, H., RÖSLER, U. and HENSEL, A., 2001. Improvement of an *invA*-based PCR for the specific detection of *Salmonella typhimurium* in organs of pigs. In: *Proceedings of the International Conference on the Epidemiology and Control of Biological, Chemical and Physical Hazards in Pigs and Pork*, 2001, Iowa. Ames: Iowa State University.
- SHANMUGASAMY, M., VELAYUTHAM, T. and RAJESWAR, J., 2011. *Inv A* gene specific PCR for detection of *Salmonella* from broilers. *Veterinary World*, vol. 4, no. 12, pp. 562. <http://dx.doi.org/10.5455/vetworld.2011.562-564>.
- SHI, C., SINGH, P., RANIERI, M.L., WIEDMANN, M. and MORENO SWITT, A.I., 2015. Molecular methods for serovar determination of *Salmonella*. *Critical Reviews in Microbiology*, vol. 41, no. 3, pp. 309-325. <http://dx.doi.org/10.3109/1040841X.2013.837862>. PMID:24228625.
- SIALA, M., BARBANA, A., SMAOUI, S., HACHICHA, S., MAROUANE, C., KAMMOUN, S., GDOURA, R. and MESSADI-AKROUT, F., 2017. Screening and detecting salmonella in different food matrices in southern Tunisia using a combined enrichment/real-time PCR method: correlation with conventional culture method. *Frontiers in Microbiology*, vol. 8, pp. 2416. <http://dx.doi.org/10.3389/fmicb.2017.02416>. PMID:29270157.
- SIBANDA, T.Z., KOLAKSHYAPATI, M., WELCH, M., SCHNEIDER, D., BOSHOFF, J. and RUHNKE, I., 2020. Managing free-range laying

- hens—Part A: frequent and non-frequent range users differ in laying performance but not egg quality. *Animals (Basel)*, vol. 10, no. 6, pp. 991. <http://dx.doi.org/10.3390/ani10060991>. PMID:32517207.
- SOUSA, A.M. and PEREIRA, M.O., 2013. A prospect of current microbial diagnosis methods. In: A. MENDEZ-VILAS, ed. *Microbial pathogens and strategies for combating them: science, technology and education*. Badajoz: Formatex Research Center.
- STRING [online], 2022 [viewed 23 April 2022]. Available from: <https://string-db.org/>
- SUÁREZ, M. and RÜSSMANN, H., 1998. Molecular mechanisms of *Salmonella* invasion: the type III secretion system of the pathogenicity island 1. *International Microbiology*, vol. 1, no. 3, pp. 197-204. PMID:10943360.
- VAEZ, H., GHANBARI, F., SAHEBKAR, A. and KHADEMI, F., 2020. Antibiotic resistance profiles of *Salmonella* serotypes isolated from animals in Iran: a meta-analysis. *Iranian Journal of Veterinary Research*, vol. 21, no. 3, pp. 188-197. <http://dx.doi.org/10.22099/ijvr.2020.36252.5296>. PMID:33178296.
- WALTMAN, W., GAST, R.K. and MALLINSON, E.T., 2008. *A laboratory manual for the isolation and identification of avian pathogens*. Ithaca: American Association of Avian Pathologists.
- WORRALL, L.J., VUCKOVIC, M. and STRYNADKA, N.C.J., 2010. Crystal structure of the C-terminal domain of the *Salmonella* type III secretion system export apparatus protein InvA. *Protein Science*, vol. 19, no. 5, pp. 1091-1096. <http://dx.doi.org/10.1002/pro.382>. PMID:20306492.
- ZHAO, X., GAO, Y., YE, C., YANG, L., WANG, T. and CHANG, W., 2016. Prevalence and characteristics of *Salmonella* isolated from free-range chickens in Shandong Province, China. *BioMed Research International*, vol. 2016, pp. 8183931. <http://dx.doi.org/10.1155/2016/8183931>. PMID:27800493.

Supplementary Material

Supplementary material accompanies this paper.

Table S1. The *invA* primer was blasted against the nucleotide database of the EBI (BLASTN, version 2.9.0+). This material is available as part of the online article from <https://doi.org/10.1590/1519-6984.263363>